

# Miralys™ Protocol Starter Kit: Immunology Version

### **Purpose**

The Starter Kit was created to provide researchers with many of the materials needed to establish the Miralys™ Laboratory Workflow in their labs. Additionally, practice slides and probe mix enable initial Miralys™ runs without using up precious samples. This kit is to be used in conjunction with MALDI HiPLEX-IHC MIRALYS™ IMAGING LABORATORY WORKFLOW document ("Miralys™ Protocol") which has been provided. If you do not have a copy, please contact support@ambergen.com to obtain one.

#### Contents

- 1. One (1) 6-plex Miralys<sup>™</sup> probe mixture store at -20°C and protect from prolonged light exposure. Details in table below.
- 2. Set of FFPE human tonsil slides store at 4°C or at room temperature
  - a. One (1) "pre-stained" slide stained with the same 6-plex Miralys™ probe mixture detailed in item 1
  - b. Two (2) "unstained" slides sourced from the same tissue block as item (a)
- 3. One (1) multi-channel digital timer for keeping time while working through the Miralys™ Protocol.
- 4. One (1) hydrophobic PAP barrier pen used in Step 8 of the Miralys™ Protocol
- 5. Four (4) spin filter units (0.45 μm) used in Step 10 of the Miralys™ Protocol
- 6. One (1) UV Lightbox with US and European power cords used in Step 16 of the Miralys™ Protocol
- 7. Two (2) pieces of filter paper used in Step 18 of the Miralys™ Protocol
- 8. One (1) glass petri dish (bottom only) used in Step 18 of the Miralys™ Protocol
- 9. One (1) plastic petri dish top only used in Step 18 of the Miralys™ Protocol
- 10. Four (4) neodymium magnets used in Step 18 of the Miralys™ Protocol



#### **Probe Mix Details**

Target	PC-MT (Da)*	Host	Reactivity	Final Concentration
CD3ε	1161.65	Rabbit	Н	2.5 μg / mL
CD68 <sub>GFAP</sub>	12461755	Rabbit	м, кН	2.5 kg5 kkg/mL
Collagen-1A1	1234.87	Rabbit	M, H	2.5 μg / mL
Ki67 例路中(Myelin Basic Protein)	13295753	Rabbit	M; R;H	1.25 kg pret / mL
PanCK	1628.78	Rabbit	M, R, H, Mk	2.5 μg / mL
VIM (Misshell Misshell Harris Profest)	1 <b>236</b> 58 <b>7</b> 8	Rabbit	MM;rR;rH, Mk	1:25   <b>½</b>
*PC-MINE Monoisotopic (M+H)+ C	of the massible forter	Rabbit	M; R; H	12.3 Hg/ME
NF-L (Neurofilament Light)	1345.74	Rabbit	M, R, H	1.25 μg / mL
SYN-I (Synapsin I)	In <del>ŝt</del> ŝu <i>c</i> tio	r <sup>Rgb</sup> för Use	M, R, H	1.25 μg / mL

### **Image the Pre-Stained Slide**

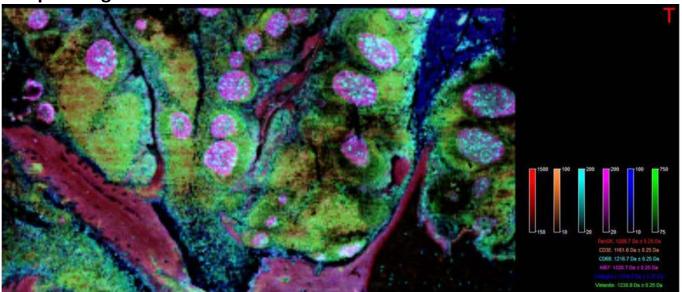
- Open the pink bubble-wrap slide envelope and the vacuum-sealed package within. Remove the pre-stained slide and re-store the unstained slides. Important note: If slides were stored cold, equilibrate to room temperature before opening the vacuum-sealed pouch.
- 2. Begin at Step 16 of the Miralys™ Protocol (photocleavage) and follow the protocol through to the end to prepare slide
- 3. Image in any MSI instrument

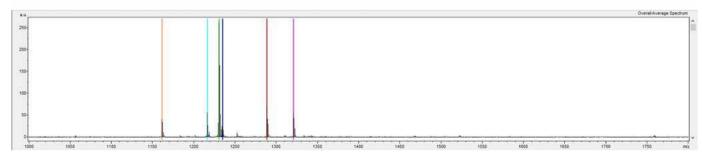
## Stain and Image the Unstained Slides

- 1. To prepare the unstained slides, begin by pre-melting as per Step 1 of the Miralys™ Protocol.
- 2. Complete Steps 2, 4, and 5 of the Miralys™ Protocol. Step 3 can be omitted because the tissue is FFPE.
- 3. Follow Steps 6 thru 8 of the Miralys™ Protocol, using alkaline antigen retrieval buffer in Step 6.
- 4. Because the Miralys™ probe is pre-mixed, perform the following in place of Step 9:
  - a. Prior to opening probe:
    - i. Vortex for 30 seconds with a benchtop vortex
    - ii. Centrifuge for 1 minute at full speed
  - b. For each slide, aliquot 24  $\mu$ L of the supplied 48  $\mu$ L probe mixture into a separate microcentrifuge tube and dilute to 400  $\mu$ L final volume with the Tissue Blocking Buffer. Vortex for 30 seconds and centrifuge again for 1 minute.
- 5. Begin again with the Miralys™ Protocol at Step 10 and follow through to the end to prepare slides.
- 6. Image in any MSI instrument



# **Sample Images**





**Top:** Example Miralys<sup>TM</sup> Results from Pre-Stained FFPE Human Tonsil Tissue Slide. MALDI mass spectrometry imaging was performed on a Bruker rapifleX<sup>TM</sup> Tissuetyper with 20  $\mu$ m spatial resolution. The image was generated from flexImaging with TIC normalization and the display intensity scale in absolute units.

**Bottom:** Mean Spectrum from the Entire Region Shown. Color-coded bars indicate the reporter peaks from the photocleaved mass-tags (bars are color-coded according to the key provided in the image in the top panel).



